



Robert Bugg

▲ A close up of toothpick weed (*A. visnaga*) covered with beneficial insects.

◀ Entomologist Andrew Corbett sprays a rubidium solution onto toothpick weed (*A. visnaga*) at Lindell Farms in Fresno County to track the movement of beneficial insects.

# Beneficial insects move from flowering plants to nearby crops

Rachael Freeman Long □ Andrew Corbett □ Celia Lamb  
Chris Reberg-Horton □ Jeff Chandler □ Michael Stimmann

**Marking studies demonstrated that lady beetles, lacewings, syrphid flies and parasitic wasps fed on nectar or pollen provided by borders of flowering plants around farms; many insects moved 250 feet into adjacent field crops. Studies using the elemental marker rubidium also showed that syrphid flies, parasitic wasps and lacewings fed on flowering cover crops in orchards and that some moved 6 feet high in the tree canopy and 100 feet away from the treated area. The use of nectar or pollen by beneficial insects helps them survive and reproduce. Therefore, planting flowering plants and perennial grasses around farms may lead to better biological control of pests in nearby crops.**

California farmers are planting strips or borders of flowering plants and perennial grasses, termed *insectary plantings*, around their farms to attract beneficial insects and thus get better biological control of pests in their crops. The idea behind these plantings is that many adult beneficial insects feed on nectar or pollen as a sole food source or to supplement their diet during periods of prey scarcity. These critical nutrients help them survive and increase their egg-laying ability. To the extent that these food resources lead to more beneficial insects around farms, insectary plantings may result in greater biological pest control.

While these plantings seem like a good idea, there are few data to support their application as an effective biocontrol tool. The objective of this study was to determine if beneficial

insects feed on insectary plants and then move into associated crops.

## Insectary plants

Insectary plants are those that produce nectar or pollen and attract beneficial insects. These include perennial and annual plants (tables 1 and 2) and perennial grasses such as deergrass (*Muhlenbergia rigens*), purple needlegrass (*Nassella pulchra*), blue wild rye (*Elymus glaucus*), meadow barley (*Hordeum brachyantherum*), California brome (*Bromus carinatus*) and Yolo slender wheatgrass (*Elymus trachycaulus majus*). Insectary plantings that contain a variety of plants with different flowering periods provide a year-round food supply for beneficial insects.

## Using rubidium to track insects

Rubidium (Rb) was used to document the feeding by beneficial insects at insectary plantings and their movement into adjacent crops. This element substitutes for the potassium in plant and insect tissues. Tests have shown that at low concentrations rubidium does not have any detectable effects on longevity or behavior of most insects (Stimmann et al. 1973). It can also be easily tracked in biological systems through the use of a flame emission spectrophotometer.

Beneficial insects were labeled by injecting or spraying solutions of rubidium chloride (RbCl) into or on in-

Fred Thomas



At the almond orchard site, lacewings, syrphid flies and parasitic wasps moved up to 100 feet away from the insectary plantings.

sectary plants. The insects obtained the label through feeding on Rb-rich nectar and pollen sources that concentrated in the plant. Because Rb occurs naturally in plant and insect tissues at fairly low and uniform levels, relatively small increases in Rb result in a measurable label at levels above the natural background concentration normally found in insects (Berry et al. 1972).

### Three field sites

From 1994 to 1995, we conducted studies at three field sites where insectary plantings are being used by growers. The first site was at Hedgerow Farms in Yolo County, with fourteen 10-year-old California lilac (*Ceanothus* 'Ray Hartman') bushes located directly across from a wheat field. The second site was at Lindell Farms in

Fresno County, with a 2-year-old, 20-foot-wide-by-660-foot-long hedgerow of perennials (table 1) and annual toothpick weed (*Ammi visnaga*). This insectary planting was surrounded by organic vegetables including beans, corn and tomatoes, and a small orchard of peaches and walnuts. The third site was in an almond orchard in Merced County, where a grower planted a mix of winter insectary annuals in 1 out of every 10 tree rows (table 2).

### Rubidium applied to plants

We injected RbCl into the California lilac plants at Hedgerow Farms. This was done once before bloom, by drilling one or two holes into each main branch at the base of the plant and injecting 7 milliliters of RbCl solution (at a very high concentration of 0.25 gm

per mL) into each hole with a syringe. Injections increased the Rb content of plants by about 10 times the naturally occurring background levels. The Lindell Farms site was sprayed with RbCl at 3,000 ppm about every 2 weeks from June until August. At the almond orchard site, four cover crop strips (6 ft by 1,000 ft) were sprayed with RbCl at 3,000 ppm about every 2 weeks from May to July. RbCl sprays increase the Rb content by 200 times the naturally occurring background levels.

### Sampling beneficial insects

Following RbCl applications, we collected beneficial insects from the insectary plants and the associated crop using 6-inch-by-12-inch sticky cards of similar color to standard yellow white-fly traps. Only those beneficial insects believed important to pest management in the crops were collected for Rb analysis (table 3).

At Hedgerow Farms, we suspended 8 traps in the canopy of individual California lilac shrubs. In the adjacent wheat, we stapled 20 traps onto wooden stakes at the level of the crop at distances of 50 feet, 150 feet, 250 feet and 450 feet from the California lilac. We changed all of the traps every 2 weeks from April until May.

At Lindell Farms, we placed 10 traps on wooden stakes every 50 feet, adjacent to and just below the height of the insectary plants. In the adjacent vegetable crops, 20 traps were stapled onto wooden stakes below the level of the crop at distances of 20 feet and 250 feet from the insectary plants. In the adjacent orchard, 20 traps were placed 6 feet high in individual trees at 50-foot intervals, from 20 feet to 250 feet away from the hedgerow. Traps were changed every 2 weeks from June to August. We collected *Hyposoter* wasps using a handheld aspirator, as we did not find them on sticky traps.

At the almond orchard site, we placed 36 traps 6 feet high in individual trees (24 ft by 24 ft spacing) at 1, 3 and 5 tree rows from the insectary mix strips. Traps were changed every 2 weeks from May until July.

We removed the beneficial insects from the traps, then identified and analyzed them for Rb content. Similar

TABLE 1. Perennial insectary plants with flowering periods designated by shaded areas

Common Name	Taxon	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sept	Oct	Nov	Dec
Willow*	<i>Salix spp.</i>												
California lilac	<i>Ceanothus spp.</i>												
Mule fat	<i>Baccharis viminea</i>												
Yarrow	<i>Achillea millefolium</i>												
Coffeeberry	<i>Rhamnus californica</i>												
Hollyleaf cherry	<i>Prunus ilicifolia</i>												
Soapbark tree	<i>Quillaja saponaria</i>												
California buckwheat	<i>Eriogonum fasciculatum</i>												
St. Catherine's lace	<i>Eriogonum giganteum</i>												
Elderberry	<i>Sambucus mexicana</i>												
Toyon	<i>Heteromeles arbutifolia</i>												
Creeping boobiella	<i>Myoporum parvifolium</i>												
Milkweed	<i>Asclepias fascicularis</i>												
Coyote brush	<i>Baccharis pilularis</i>												

\*Highlighted plants are those found in the hedgerow at Lindell Farms (Bugg et al. 1998)



beneficial insect species were collected from "control" fields, located more than 2 miles away from treated hedgerows, to determine the naturally occurring background Rb content.

*Trichogramma* controls were obtained from a laboratory colony. These data provided the criteria for deciding if an individual beneficial insect captured within the insectary plantings and associated crops was labeled (table 4).

### Analysis of samples

Insects were chemically digested and analyzed for Rb content using standard methodologies (Berry et al. 1972). Insects were considered labeled if their Rb concentration was more than three standard deviations greater than the control means. In this way, the probability of incorrectly identifying an insect as labeled with Rb is very low (less than 1%).

### Labeling of beneficial insects

**Hedgerow Farms.** In the California lilac, 67% (sample size of n = 15) of the green lacewings, and 40% (n = 10) of the lady beetles were labeled with Rb, following injections with RbCl. In the adjacent wheat, 2% of the green lacewings were labeled at 50 feet and 2% were labeled at 450 feet (n = 47). We did not find any labeled lady beetles in the adjacent wheat.

**Lindell Farms.** RbCl sprays in the insectary hedgerow also resulted in marking of beneficial insects (fig. 1). In the insectary plantings, 61% of the green lacewings were labeled, plus 20% of the *Trichogramma*, and 55% of the lady beetles. We did not find any labeled *Hyposoter* wasps in the insectary

plantings, even though these wasps are known to feed on nectar of flowering plants. We suspect that these wasps were visiting the plants in a manner that escaped our detection, possibly due to the time of day or their foraging behavior.

In the adjacent vegetables (fig. 1), 24% of the green lacewings were labeled 20 feet from the hedgerow, but none were labeled at 250 feet. For lady beetles, 27% were labeled at 20 feet and 23% were labeled at 250 feet from the hedgerow. For *Hyposoter* wasps, 17% were labeled at 20 feet and 47% were labeled at 250 feet from the hedgerow. None of the *Trichogramma* were labeled in the nearby crops.

In the adjacent orchard, 10% of the green lacewings were labeled at 20 feet (n = 31) and 33% (n = 3) were labeled at 150 feet from the hedgerow. No labeled green lacewings were found in the trees at any other distances (n < 3 for all these samples). Ten percent of the lady beetles (n = 10) and 2.5% (n = 40) of the *Trichogramma* were labeled at 20 feet from the hedgerow.

**Almond orchard site.** RbCl marking of the annual insectary mix also showed labeling of beneficial insects (fig. 2). In the trees, 27% of the green

lacewings were labeled adjacent to the insectary mix, but none were labeled further away. For brown lacewings, 39% were labeled next to the insectary mix, 28% were labeled at 50 feet and 17% were labeled at 100 feet. For syrphid flies, 81% were labeled adjacent to the insectary mix, 74% were labeled at 50 feet, and 70% were labeled at 100 feet. For *Macrocentrus* wasps, 8.1% were labeled next to the insectary mix, 4.3% were labeled at 50 feet, and 4.5% were labeled at 100 feet.

### Underestimation of numbers

Although the percentage of labeled beneficial insects was low in some samples, these data probably underestimate the numbers of beneficial in-

TABLE 2. Winter annual insectary plants at the almond orchard

Common Name	Taxon
Subterranean clover	<i>Trifolium subterraneum</i>
Common vetch	<i>Vicia sativa</i>
White sweetclover	<i>Trifolium repens</i>
Crimson clover	<i>Trifolium incarnatum</i>
Rye	<i>Secale cereale</i>
Triticale	<i>X Triticosecale</i>
Barley	<i>Hordeum vulgare</i>
Bee phacelia	<i>Phacelia tanacetifolia</i>
Coriander	<i>Coriandrum sativum</i>
Sweet alyssum	<i>Lobularia maritima</i>
Tidy tips	<i>Layia platyglossa</i>
Celery	<i>Apium graveolens</i>

TABLE 3. Dominant beneficial insects collected and analyzed for rubidium content at our study sites

Beneficial insect	Taxon	Primary prey
Green lacewings	<i>Chrysoperla carnea</i>	Aphids
Brown lacewings	<i>Hemerobius</i> sp.	Aphids
Lady beetles	<i>Hippodamia convergens</i>	Aphids
Syrphid flies	<i>Toxomerus</i> sp.	Aphids
Parasitic wasps	<i>Hyposoter</i> sp.	Caterpillars
	<i>Trichogramma</i> sp.	Moth eggs
	<i>Macrocentrus</i> sp.	Peach twig borers
		Leafrollers

TABLE 4. Highest rubidium level found in beneficial insects in crops versus naturally occurring background levels (controls)

Beneficial insects	Crops			Control			Source of controls
	3 highest Rb values*			Mean	Standard deviation	Sample size	
	nanograms of Rb per insect						
Green lacewings	2,000	1,200	1,000	89	58	94	Wheat
Brown lacewings	486	369	255	18	13	18†	Wheat
Lady beetles	1,000	940	900	64	36	220	Wheat
<i>Hyposoter</i> wasps	330	300	280	15	14	34	Sugarbeet
Syrphid flies	1,240	1,240	600	14	5	24‡	Almonds
<i>Macrocentrus</i> wasps	43	28	28	3	1	67	Almonds
<i>Trichogramma</i> wasps	1.6	1.4	0.7	0.1	0.15	26§	Lab-reared colony

\*Insects were considered labeled if their Rb concentration was more than 3 standard deviations from the control means. For normally distributed populations this means that P < 0.001. For sample sizes less than 30, the following standard deviations were used: †4.0, ‡3.8 and §3.7.





Hedgerow of insectary shrubs and native grasses, Yolo County.



Scientists found that 70% of the syrphid flies in almond trees were marked with rubidium 100 feet away from the insectary planting.

sects that are using the insectary plants. Our laboratory studies showed that green lacewings need to feed on Rb-marked floral resources for up to 24 hours to acquire detectable levels of Rb. If beneficial insects are feeding on both marked and unmarked plants in the field, the impact of the Rb label will likely be diluted. For example, at Hedgerow Farms, where we had low levels of labeling, there is an abundance of floral resources other than

our Rb-marked California lilac for beneficial insects to feed on. In addition, our laboratory studies showed that green lacewings excrete the label within 24 hours.

The results of this study demonstrate that beneficial insects feed on nectar or pollen provided by insectary plants, and that they move into associated crops. Because many Rb-labeled individuals were trapped 250 feet from the insectary plants, our data

suggest strips or borders of flowering plants may help beneficial insects — even at long distances. The survival of many beneficial insect species depends on nectar or pollen so adding insectary plantings to the agricultural landscape may enlarge their populations and increase their activity in crops.

R. Freeman Long is UC Cooperative Extension Farm Advisor, Yolo and Solano counties; A. Corbett is Postdoctoral Researcher, Entomology, UC Davis; C. Lamb is Field Assistant, Agronomy, UC Davis; C. Reberg-Horton is Graduate Student, Agronomy, UC Davis; J. Chandler is Consultant, Corn Flower Farms, Elk Grove; and M. Stimmann is Extension Environmental Toxicologist, UC Davis.

This research was supported by the California Tomato Research Institute and the U.S. Environmental Protection Agency.

## References

- Berry WL, Stimmann MW, Wolf WW. 1972. Marking of phytophagous native insects with rubidium: A proposed technique. *Annals Entomol* 65:236-8.
- Stimmann MW, Wolf WW, Berry WL. 1973. Cabbage loopers: The biological effects of rubidium in the larval diet. *J Econ Entomol* 66:324-6.
- Bugg RL, Anderson JH, Thomsen CD, Chandler J. 1998. *Farmscaping: Restoring native biodiversity to agricultural settings*. In Pickett CH and Bugg RL (Eds.) *Enhancing biological control: habitat management to promote natural enemies of agricultural pests*. University of California Press, Berkeley, CA. (In press).

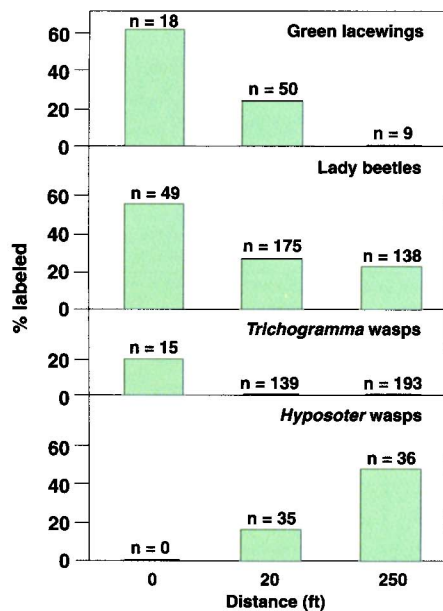


Fig. 1. Percent Rb-labeled beneficial insects in the insectary hedgerow (0 ft) and in the adjacent vegetable crop at 20 ft and 250 ft (n = total number of insects captured).

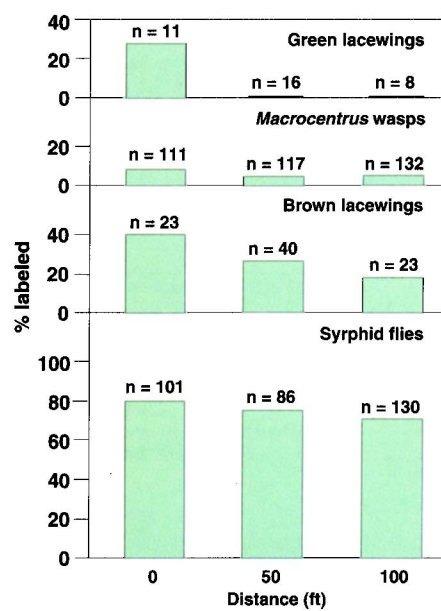


Fig. 2. Percent Rb-labeled beneficial insects trapped in the almond trees at 0, 50 and 100 ft from the Rb-labeled insectary cover crop mix (n = total number of insects captured).